

3.10: Mitochondrial Calcium Ion Transport

Influx

Mitochondria isolated from various types of animal cells—but, interestingly, not those from plant cells—can rapidly accumulate exogenous Ca^{2+} .⁵⁹ The transporter is located in the inner membrane and the driving force behind the Ca^{2+} transport appears to be merely the high potential difference across this membrane ($\Delta\Psi \approx 150$ to 180 mV, negative in the inner matrix). This potential difference is fairly closely maintained by the pumping out of H^+ from the matrix by cell respiration. For the transport of 1 mol Ca^{2+} from the "outside" (= cytoplasm) to the "inside" (= inner mitochondrial matrix), we may deduce from Equation (3.4) that the free-energy change ΔG may be written ($\Delta n_{\text{Ca}^{2+}} = -1$)

$$\Delta G = -RT \cdot \ln \frac{[\text{Ca}^{2+}]_o}{[\text{Ca}^{2+}]_i} - 2F\Delta\Psi. \quad (3.11)$$

From this analysis it may be inferred that the limiting Ca^{2+} concentration (or activity) ratio that can be achieved by this **electrogenic pump** (i.e., $\Delta G = 0$) is

$$\frac{[\text{Ca}^{2+}]_o}{[\text{Ca}^{2+}]_i} = e^{\frac{-2F\Delta\Psi}{RT}} \quad (3.12)$$

With $\Delta\Psi = 150$ mV, this ratio is calculated to be 8.4×10^{-6} at 25°C . It is evident that, as long as the Ca^{2+} influx would not lower the membrane potential difference, the Ca^{2+} **uniporter** has a very high pumping potential. Measured values of the pumping rate, V_{max} , are indeed high (>10 nmol/mg protein⁵⁹) and probably limited only by the rate of electron transport and H^+ extrusion in the mitochondria.

Mitochondria may accumulate large quantities of Ca^{2+} , probably to maintain electroneutrality. To prevent the buildup of high concentrations of free Ca^{2+} (and of osmotic pressure), phosphate ions are also transported into the inner matrix, where an amorphous calcium phosphate—or possibly a phosphocitrate⁶⁰—is formed. The equilibrium concentration of free Ca^{2+} in the mitochondrial matrix may as a result be comparatively low, on the order of $1 \mu\text{M}$.

The molecular nature of the mitochondrial Ca^{2+} uniporter continues to be elusive, and needs to be studied further.

Efflux

Mitochondria, as well as SR, release Ca^{2+} ions by mechanisms other than "back leakage" through the pumps. In mitochondria from excitable cells, the efflux occurs mainly through an antiport, where 2Na^+ ions are transported inward for every Ca^{2+} ion departing for the cytosolic compartment.⁶¹ In other cells there is evidence for the dominance of a 2H^+ - Ca^{2+} antiport.⁵⁹ In all likelihood the Ca^{2+} efflux is regulated, possibly by the redox state of pyridine nucleotides in the mitochondria. As with the Ca^{2+} uniporter, few details on the molecular nature of the antiporters are presently available.

Ca^{2+} Efflux from Non-mitochondrial Stores

Release of Ca^{2+} from ER and SR presently appears to be the prime effect of the new intracellular messenger 1,4,5-triphosphoinositol ($1,4,5\text{-IP}_3$) released into the cytoplasm as a result of an external hormonal stimulus (see Section IV.C). It seems that receptors for $1,4,5\text{-IP}_3$ have been established on ER, and that the binding of $1,4,5\text{-IP}_3$ causes a release of Ca^{2+} stored in this organelle.^{62,63,170,171} In addition to the receptor-controlled Ca^{2+} efflux, there may be other pathways for Ca^{2+} release, and Ca^{2+} mobilization may be regulated by other intracellular entities, the Ca^{2+} ions themselves included.

Other Voltage-gated or Receptor-activated Ca^{2+} Channels

In addition to the transport pathways already discussed, some cells seem to have Ca^{2+} channels in the plasma membrane that can be opened by the action of an agonist on a receptor or that are gated in response to changes in membrane potential.⁶⁴ For example, Ca^{2+} channels can be opened by nicotinic cholinergic agonists⁶⁵ or by the excitatory amino acid N-methyl-D-aspartate (NMDA).⁶⁶ Endocrine cells and also some muscle and neuronal cells have voltage-sensitive Ca^{2+} channels.^{67,68} We will not discuss these further, but merely point to their existence. We finally note that during the last few years knowledge about the mechanisms of Ca^{2+} entry and release to and from extracellular and intracellular pools has increased dramatically, and we refer the reader to recent reviews of the field.^{175,176}

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